

## IN THE SPECIFICATION

After the Abstract on page 44, insert new Sequence Listing pages 1-12 submitted herewith.

Replace the paragraph beginning with "Thus, the present invention relates" on page 3 of the specification with the following paragraph.

Thus, the present invention relates to a method for the identification of hetero-associating (poly)peptides comprising the steps of:

(a) providing a library A of (poly)peptides/proteins comprising (poly)peptides  $A_m$  having the general formula:

VAQLXEXVKTLXAXZYELXSXVQRLXEXVAQL (SEQ ID NO. 1)

wherein X represents a mixture of E, K, Q, and R, and wherein Z represents a mixture of N and V,

(b) providing a library B of (poly)peptides/proteins comprising (poly)peptides  $B_n$  having the general formula:

VDELXAXVDQLXDXZYALXTXVAQLXKXVEKL (SEQ ID NO. 2)

wherein X represents a mixture of E, K, Q, and R, and wherein Z represents a mixture of N and V;

(c) combining in a common medium the (poly)peptides/proteins of said libraries A and B; and

(d) screening or selecting for a screenable or selectable property caused by the hetero-association of a (poly)peptide  $A_m$  with a (poly)peptide  $B_n$ .

Replace the paragraph beginning "In another embodiment" on page 5 of the specification with the following paragraph.

In another embodiment, the present invention relates to a hetero-associating (poly)peptide  $A_m$  taken from the list of:

WINZIPA1: VAQLEEKVKTLRAQNYELKSRVQRLREQVAQL (SEQ ID NO. 3)

WINZIPA2: VAQLRERVKTLRAQNYELESEVQRLREQVAQL (SEQ ID NO. 4)

WINZIPA3: VAQLQEKVKTLRARNYELKSEVQRLQEEKVAQL (SEQ ID NO. 5)

WINZIPA4: VAQLEEQVKTLQARNYELKSKVQRLKEKVAQL (SEQ ID NO. 6)

WINZIPA5: VAQLEERVKTLRAQNYELKSKVQRLQEEQVAQL (SEQ ID NO. 7)

WINZIPA6: VAQLEEQVKTLQARNYELKSKVQRLRERVAQL (SEQ ID NO. 8)

WINZIPA7: VAQLQEQVKTLQARNYELESEVQRLKEQVAQL (SEQ ID NO. 9)

WINZIPA8: VAQLEERVKTLQARNYELESEVQRLKERRVAQL (SEQ ID NO. 10)

WINZIPA9: VAQLEEKVKTLKARNYELKSKVQRLKEKVAQL (SEQ ID NO. 11)

WINZIPA10: VAQLQEEVKTLQAENYELRSEVQRLEEEVAQL (SEQ ID NO. 12)  
WINZIPA11: VAQLRERVKTLRARNYELQSKVQRLKERVEAQL (SEQ ID NO. 13)

Replace the paragraph beginning “Furthermore, the present invention” on page 6 of the specification with the following paragraph.

Furthermore, the present invention relates to a hetero-associating (poly)peptide B<sub>n</sub> taken from the list of:

WINZIPB1: VDELQAEVDQLQDENYALKTKVAQLRKKVEKL (SEQ ID NO. 14)  
WINZIPB2: VDELKAEVDQLQDQNYALRTKVAQLRKEVEKL (SEQ ID NO. 15)  
WINZIPB3: VDELEAEVDQLKDQNYALKTKVAQLQKQVEKL (SEQ ID NO. 16)  
WINZIPB4: VDELRAKVDQLQDENYALETEVAQLQKRVEKL (SEQ ID NO. 17)  
WINZIPB5: VDELEAEVDQLEDQNYALQTRVAQLEKRVEKL (SEQ ID NO. 18)  
WINZIPB6: VDELKAKVDQLKDKNYALRTKVAQLRKKVEKL (SEQ ID NO. 19)  
WINZIPB7: VDELRAQVDQLQDKNYALRTRVAQLKKRVEKL (SEQ ID NO. 20)  
WINZIPB8: VDELQAEVDQLQDQNYALRTQVAQLKKKVEKL (SEQ ID NO. 21)  
WINZIPB9: VDELRAQVDQLEDQNYALETQVAQLEKEVEKL (SEQ ID NO. 22)  
WINZIPB10: VDELQAKVDQLKDQNYALQTKVAQLQKRVEKL (SEQ ID NO. 23)  
WINZIPB11: VDELRAEVDQLEDQNYALRTRVAQLRKQVEKL (SEQ ID NO. 24)

Replace the paragraph beginning “Trinucleotide codons (27) were used” on page 18 of the specification with the following paragraph.

Trinucleotide codons (27) were used to code for randomized positions, all other positions were made with mononucleotides.

Library A:

TACTGTGGCGCAACTGNNNGAANNNGTGAAAACCCTTNNNGC-  
TNNNXXXTATGAACTTNNNTCTNNNGTGAGCGCTTGNNNGAGNNNGT  
TGCCCAGCTTGCTA (SEQ ID NO. 25) (encoding  
VAQLXEXVKTLXAXZYLXSXV QRLXEXVAQL (SEQ ID NO. 26), wherein  
X represents a mixture of E, K, Q, and R, and wherein Z represents a mixture of N  
and V); libraryB:  
CTCCGTTGACGAACTGNNNGCTNNNGTTGACCAGCTGNNNGACNNNX  
XXTACGCTCTGNNNACCNNNGTTCGCAGCTGNNNAAANNNGTGAAAA

AGCTGTGATA (SEQ ID NO. 27) (encoding  
VDELXAXVDQLXDXZYALXTXVAQL- KXXVEKL (SEQ ID NO. 28),  
wherein X represents a mixture of E, K, Q, and R, and wherein Z represents a  
mixture of N and V) (NNN = equimolar mixture of the trinucleotides AAG, CAG,  
GAG, CGT; XXX = equimolar mixture of the trinucleotides AAT, GTT).

Replace the paragraph beginning "Generation of the second strand" on page 18 of  
the specification with the following paragraph.

Generation of the second strand and introduction of SalI and NheI restriction sites  
were achieved by PCR using the primers prA-fwd:  
GGAGTACTGGCATGCAGTCGACTACTGTGGCGCAACTG (SEQ ID NO.  
29) and prA-rev: GGACTAGTACCTTCGCTAGCAAGCTGGGCAAC (SEQ ID  
NO. 30) or prB-fwd:  
GGAGTACTGGCATGCAGTCGACCTCCGTTGACGAACTG (SEQ ID NO.  
31) and prB-rev: GGACTAGTGCTAGCTTCTGACAGCTTTTCCAC (SEQ ID  
NO. 32), respectively. This resulted in a 142 bp double-stranded oligonucleotide  
for either library.

Replace the paragraph beginning "Library A and B were both digested " on page  
18 of the specification with the following paragraph.

Library A and B were both digested with SalI and NheI, gel purified and ligated to the  
appropriate vector (Fig 2) yielding the plasmids LibA-DHFR[1], LibB-DHFR[2],  
LibB-DHFR[2:I114A] (Fig. 2A). After subcloning, the resulting linker between  
either library and DHFR fragment was: A(SGTS)<sub>2</sub> STSSGI (SEQ ID NO. 33) for  
LibA and SEA(SGTS)<sub>2</sub> STS (SEQ ID NO. 34) for LibB. To achieve maximal library  
representation, the ligation mixes were individually electroporated into XL1-Blue  
cells and selected with ampicillin on rich medium (LB). A 2- to 7-fold over-  
representation of each library was obtained. The resulting colonies were pooled and  
the plasmid DNA purified such that supercoiled plasmid DNA was obtained for  
cotransformation. The supercoiled DNA was cotransformed in BL21 cells yielding  
about  $4 \times 10^6$  double-transformants. We used BL21 cells with a transformation  
efficiency of no less than  $5 \times 10^7$  transformants per mg of DNA using 200 pg of  
DNA, or  $2 \times 10^7$  transformants per mg using 500 ng of DNA. In cotransformations,  
the occurrence of double transformation was calculated as the number of colonies  
growing under selective pressure with trimethoprim (described below) divided by the  
number growing in the absence, when cotransformed with equal amounts of each